

Certification systems for reproductive material of landscape shrubs and trees in Germany and potential possibilities for verification of their source

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International Conference of
European Seed Kilns,
Bernkastel-Kues 04 - 07 June 2013

Thünen Institute of Forest Genetics

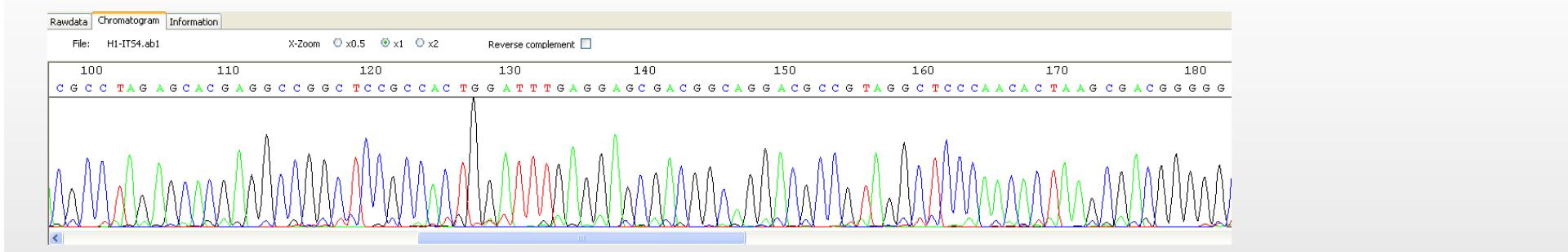
Großhansdorf and Waldsieversdorf



Federal Ministry
of Food, Agriculture
and Consumer Protection

- Provenance and breeding research
- Pathogen resistance research
- Ecological genetics
- Genome research

<http://www.ti.bund.de/>



Outline

- Current certification systems for forest and landscape reproductive materials in Germany – An overview
- Biological basics of genetic markers
- Application of markers for verification of source
- Outlook



Certification systems in Germany

Federal Nature Conservation Act 2010

Guidelines 2012
developed by a task group and the
Federal Ministry for the Environment



6 regions of provenances—
scaled by different environmental
conditions



Certification systems in Germany

Federal Nature Conservation Act 2010

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6 regions of provenances—
scaled by different environmental
conditions

- 1 Norddeutsches Tiefland
- 2 Mittel- und Ostdeutsches Tief- und Hügelland
- 3 Südostdeutsches Hügel- und Bergland
- 4 Westdeutsches Bergland und Oberrheingraben
- 5 Schwarzwald, Württembergisch-Fränkisches Hügelland
und Schwäbisch-Fränkische Alb
- 6 Alpen und Alpenvorland



Certification systems in Germany

		Field of action
ZÜF		Germany
FfV		Germany
ZgG		Germany
RAL - GZ		Germany
eab		Bavaria
pro agro		Brandenburg
EZG		Baden-Württemberg
EsB		Northern lowlands

Certification systems in Germany

		Field of action	Forest/ Landscape	Since
ZÜF		Germany	Forest	2002
FfV		Germany	Forest	2006
			Landscape	2011
ZgG		Germany	Landscape	2011
RAL - GZ		Germany	Landscape	2011
eab		Bavaria	Landscape	2001
pro agro		Brandenburg	Landscape	2002
EZG		Baden-Württemberg	Landscape	2006
EsB		Northern lowlands	Landscape	2007

Certification systems in Germany

		Field of action	Forest/ Landscape	Since	Documentation	Operating control	Plausibility
ZÜF		Germany	Forest	2002	+	+	+
FfV		Germany	Forest	2006	+	+	+
			Landscape	2011	+	+	
ZgG		Germany	Landscape	2011	+	+	(+)
RAL - GZ		Germany	Landscape	2011	+	+	+
eab		Bavaria	Landscape	2001	+	+	+
pro agro		Brandenburg	Landscape	2002	+	+	
EZG		Baden-Württemberg	Landscape	2006	+	+	+
EsB		Northern lowlands	Landscape	2007	+	+	

Certification systems in Germany

		Field of action	Forest/ Landscape	Since	Documentation	Operating control	Plausibility	Retain samples	Genetic markers
ZÜF		Germany	Forest	2002	+	+	+	+	+
FfV		Germany	Forest	2006	+	+	+	+	+
			Landscape	2011	+	+		+	
ZgG		Germany	Landscape	2011	+	+	(+)		
RAL - GZ		Germany	Landscape	2011	+	+	+		
eab		Bavaria	Landscape	2001	+	+	+		
pro agro		Brandenburg	Landscape	2002	+	+			
EZG		Baden-Württemberg	Landscape	2006	+	+	+		
EsB		Northern lowlands	Landscape	2007	+	+			

Certification systems in Germany

Current process

The task group is discussing the minimum standards for certification

Members

- Federal Ministry for the Environment, Nature Conservation and Nuclear Safety,
- Federal Ministry of Food, Agriculture and Consumer Protection,
- Federal Ministry of Transport, Building and Urban Development,
- Federal and state working group for forest genetic resources,
- Inter-trade organisations (nurseries)
- ...

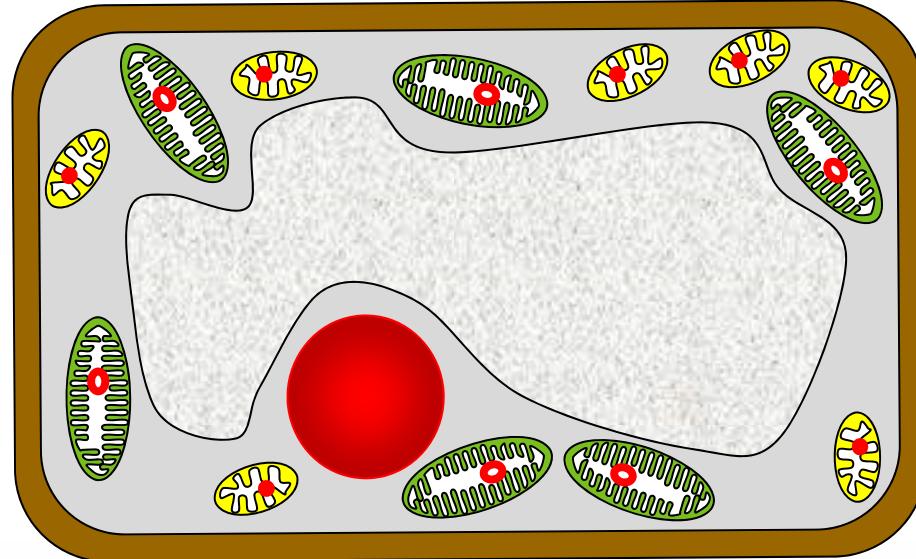
- How we can carry out genetic analyses?
- How can genetic analyses accomplish the verification of sources of reproductive material?



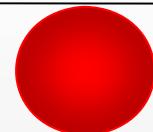
Genetic markers

Basics

Scheme of a plant cell



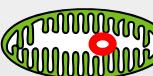
Inheritance mode



Nucleus

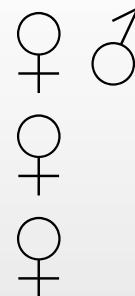


Mitochondria

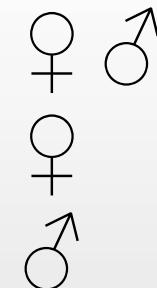


Chloroplasts

Angiosperms



Gymnosperms



Genetic markers

What is a genetic marker?

- Localisation at a certain position in genome (DNA, protein as gene product)
- Reproducible characterisation
- Independently from ontogenetic stage and environment
- Exhibits variation

Typical variation patterns

Nuclear markers

Large variation within populations,
individual identification possible

Example „**Nuclear microsatellites**“

Organelle markers

lower variation, but rather
geographic differentiation

Example „**cpDNA haplotypes**“

Genetic markers

Step by step

- Suitable sample materials: leaves, buds, seeds, root tips, cambium, ...
- Extraction of total DNA: incl. nucleus, organelles, bacteria, fungi, ...
- Amplification of selected DNA fragments by PCR (Polymerase Chain Reaction) with specific primers
- Detection of fragments by electrophoresis: capillary DNA sequencer, slab gels

Genetic markers

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- Extraction of total DNA: incl. nucleus, organelles, bacteria, fungi, ...
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for each species
and
for each marker

Genetic markers

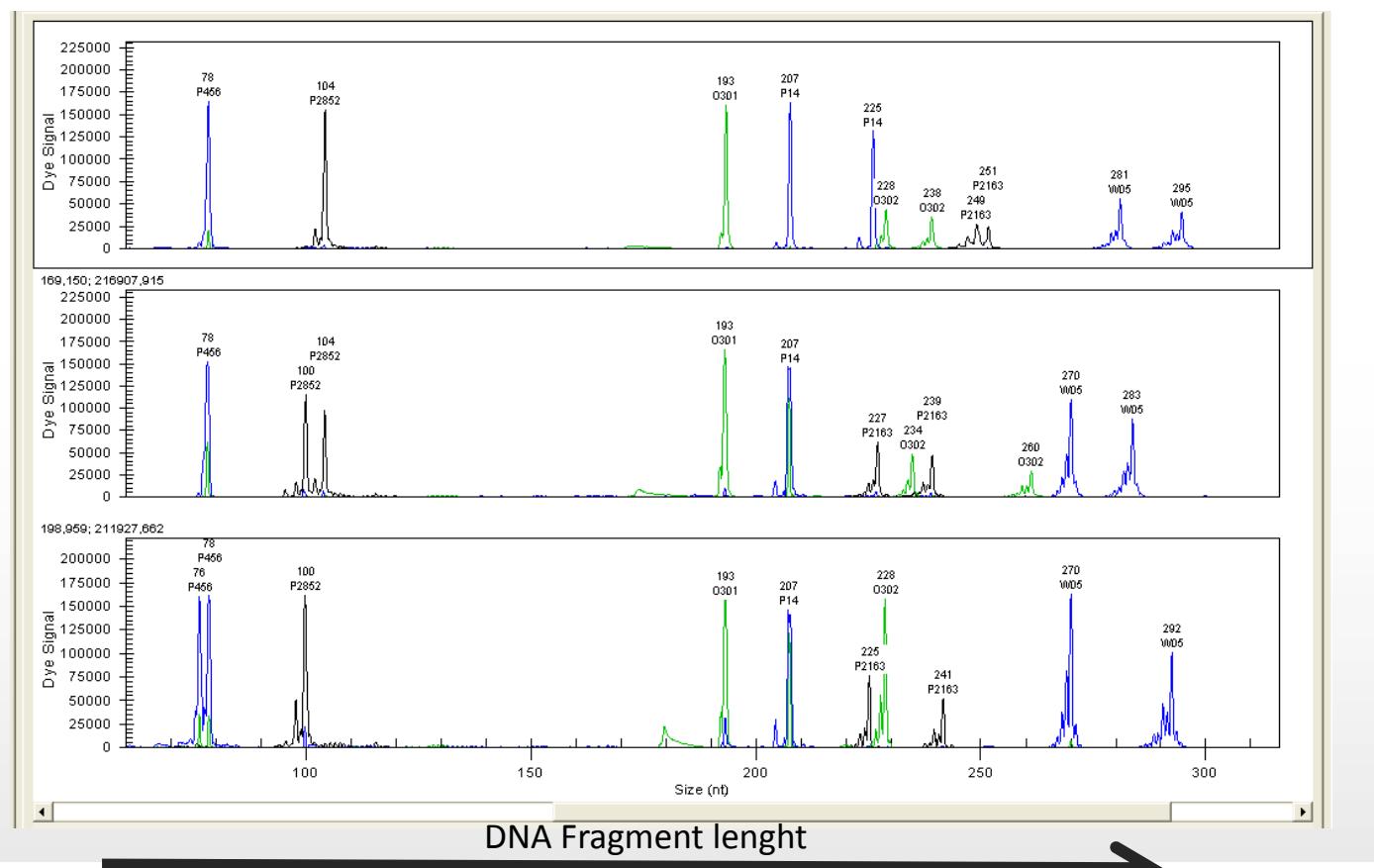
Microsatellite markers:

Variation of DNA fragment length caused by different numbers of repeats of short sequences

...ACCATGCCCTAAA**GAAGAAGAAGAAGAAGAAAGTATTGTGTATAGGTCA...**
...ACCATGCCCTAAA**GAAGAAGAAGAAGAAGAAGAAGAAGAAAGTATTGTGTATAGGTCA...**
...ACCATGCCCTAAA**GAAGAAGAAGAAGAAGAAGAAAGTATTGTGTATAGGTCA...**
...ACCATGCCCTAAA**GAAGAAGAAGAAGAAGAAGAAGAAAGTATTGTGTATAGGTCA...**
...ACCATGCCCTAAA**GAAGAAGAAGAAAGTATTGTGTATAGGTCA...**
...ACCATGCCCTAAA**GAAGAAGAAGAAGAAAGTATTGTGTATAGGTCA...**

Genetic markers

„Genetic fingerprint“ of 3 black poplars
(*Populus nigra*) at 7 microsatellite markers



Genetic markers

Number of possible combinations for individual genotypes

Number of markers	Number of alleles per marker	Number of possible combinations (Genotypes)
1	2 (A, B)	3 (AA, AB, BB)

Genetic markers

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1	2 (A, B)	3 (AA, AB, BB)
1	3 (A, B, C)	6 (AA, AB, AC, BB, BC, CC)

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1	2 (A, B)	3 (AA, AB, BB)
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1	4	10
1	5	15
1	10	55
1	15	120

Genetic markers

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1	2 (A, B)	3 (AA, AB, BB)
1	3 (A, B, C)	6 (AA, AB, AC, BB, BC, CC)
1	4	10
1	5	15
1	10	55
1	15	120
2	5	225
2	10	3 025
5	5	759 375
5	10	$5,03 * 10^9$
10	5	$5,77 * 10^{11}$
10	10	$2,53 * 10^{17}$
15	5	$4,38 * 10^{17}$

Application of markers for verification of sources

Three cases of application of markers for verification of sources

- Identity of clones
- Descent of a population
- Assignment to large-scale regions



Application of markers for verification of sources

1. Identity of clones

Comparison of a sample with a reference



Exclusion of identity,

if differences were detected



Confirmation of identity,

if no differences were detected

+

if the probability of identity by chance is small enough

Application of markers for verification of sources

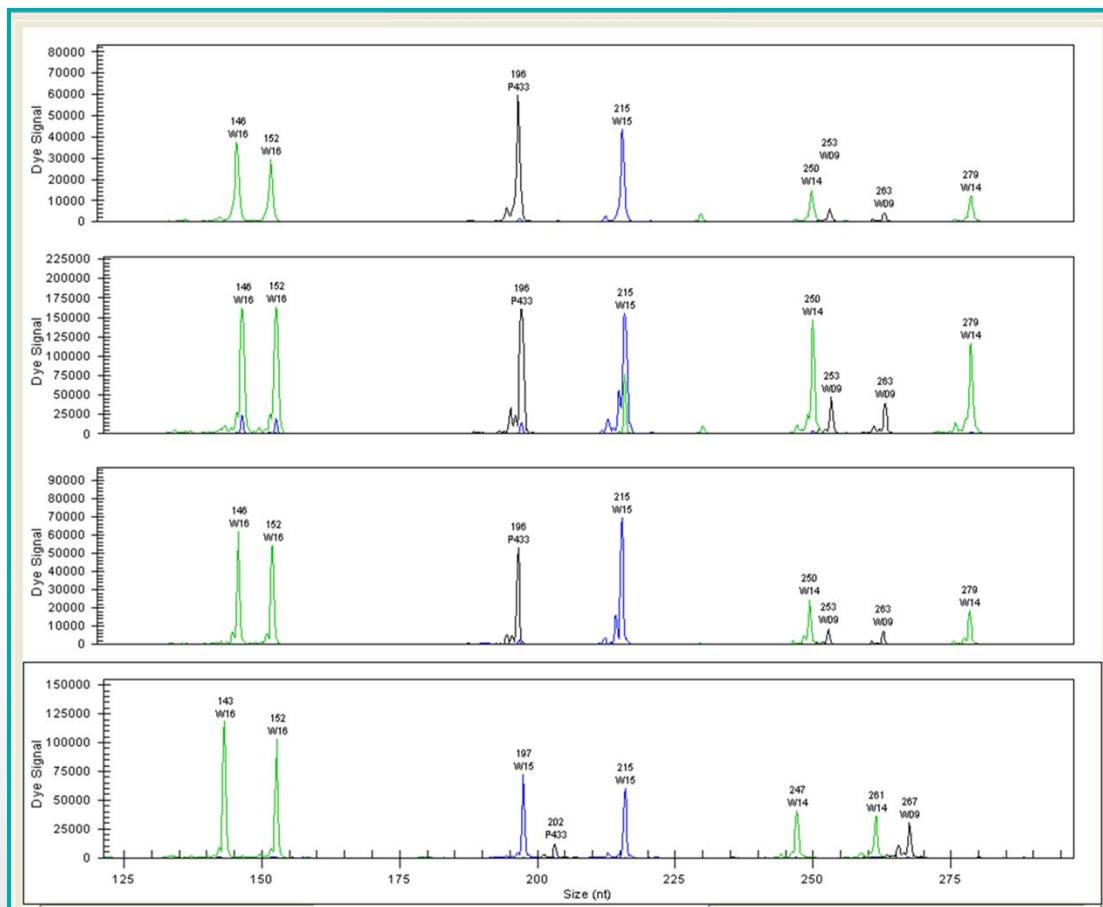
1. Identity of clones - Example

Donor plant

Cutting 1
identical

Cutting 2
identical

Cutting 3
different



Application of markers for verification of sources

2. Descent of a population

With retain samples

Comparison of seed or plant lots with retain samples,
application of exclusion principle

Without retain samples

Comparison of seed or plant lots with the parental population:

Possible for defined populations or seed orchards by exclusion principle

No assignment to a region of occurrence without retain sample or without known parental population!!

Application of markers for verification of sources

2. Descent of a population - Example

The descent of seeds from a certain tree can be excluded for individual seeds carrying genotypes without maternal alleles.

Sample	Marker005	Marker410
Seed tree	106 - 126	124 - 134
Seed 1	106 - 126	134 - 134
Seed 2	106 - 126	134 - 134
Seed 3	106 - 126	124 - 132
Seed 4	106 - 118	130 - 132
Seed 5	106 - 106	134 - 134
Seed 6	126 - 126	130 - 132
Seed 7	110 - 126	132 - 132
Seed 8	110 - 110	130 - 132

No maternal alleles

Konnert M. 2005. Erfolge (und Grenzen) bei dem Herkunfts-nachweis mittels Isoenzym- und DNA-Analysen.
In: 26. Tagung der Arbeitsgemeinschaft Forstgenetik und Forstpflanzenzüchtung 2005 in Fulda.
Hann. Münden, HESSEN-FORST, 49-57.

Application of markers for verification of sources

2. Descent of a population - Example

The descent of seeds from a certain tree can be excluded for individual seeds carrying genotypes without maternal alleles.

Sample	Marker005	Marker410	Result
Seed tree	106 - 126	124 - 134	
Seed 1	106 - 126	134 - 134	Ancestry possible
Seed 2	106 - 126	134 - 134	Ancestry possible
Seed 3	106 - 126	124 - 132	Ancestry possible
Seed 4	106 - 118	130 - 132	Ancestry excluded
Seed 5	106 - 106	134 - 134	Ancestry possible
Seed 6	126 - 126	130 - 132	Ancestry excluded
Seed 7	110 - 126	132 - 132	Ancestry excluded
Seed 8	110 - 110	130 - 132	Ancestry excluded

Application of markers for verification of sources

3. Assignment to large-scale regions

Geographic patterns

- can be observed regarding colonisation routes after ice age starting from different glacial refugia
- can result from restricted gene flow among populations (isolation by distance)

Precondition:  Genetic inventory

Application of markers for verification of sources

3. Assignment to large-scale regions

Geographic pattern of cpDNA haplotypes (PCR-RFLPs)

Example *Frangula alnus*

78 populations,
322 individuals

Hampe A, Arroyo J, Jordano P, Petit RJ. 2003. Rangewide phylogeography of a bird-dispersed Eurasian shrub: contrasting Mediterranean and temperate glacial refugia. *Molecular Ecology*, 12: 3415-3426.

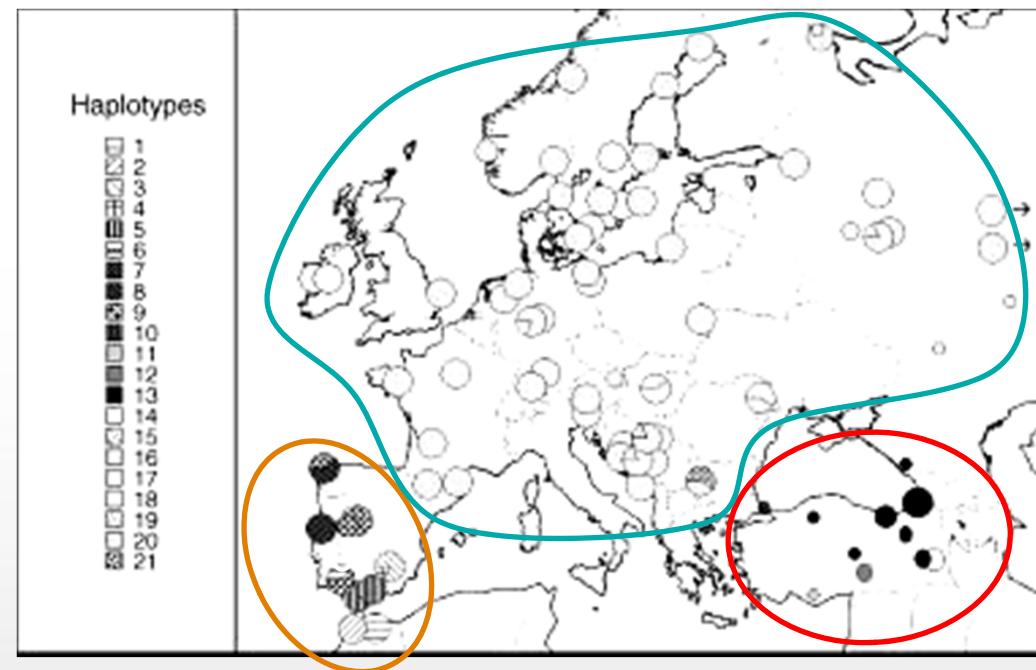


Fig. 3 Geographical distribution of *Frangula alnus* chloroplast haplotypes. Circle sizes are proportional to the number of analysed samples per population ($n = 1-5$).

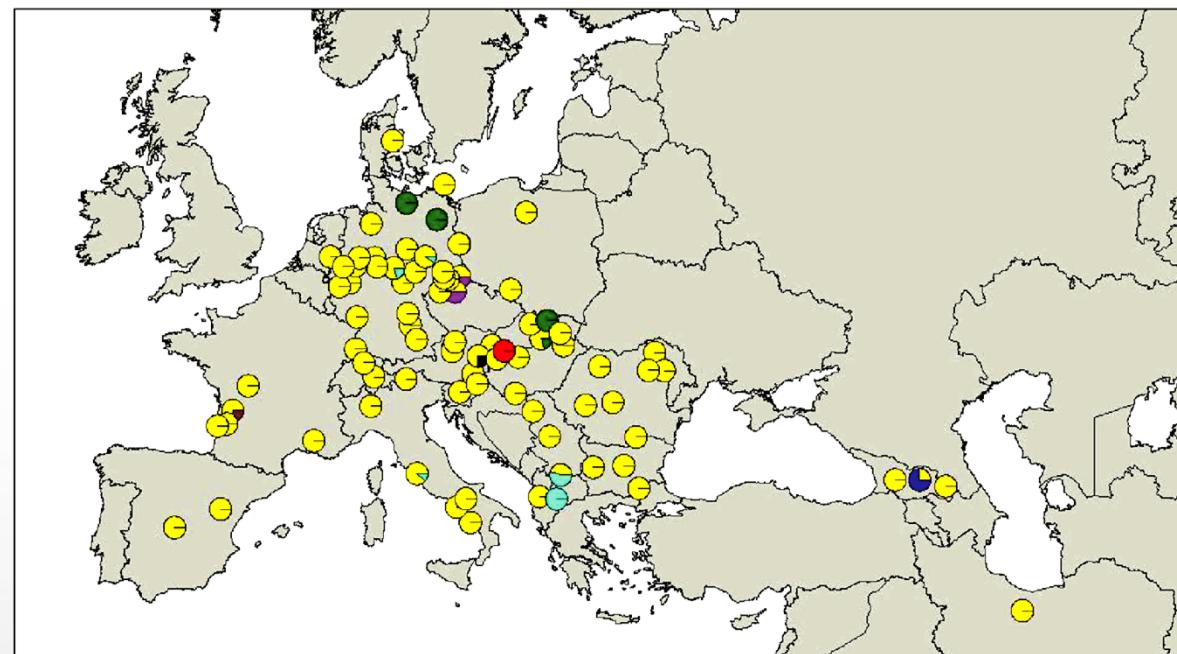
Application of markers for verification of sources

3. Assignment to large-scale regions

Geographic pattern of cpDNA haplotypes (PCR-RFLPs)

Example *Cornus sanguinea*

86 populations,
673 individuals



Liesebach & Götz, Silvae Genetica (2008) 57: 291-300

Figure 4. – Geographic distribution of eight chloroplast haplotypes identified in *Cornus sanguinea* (haplotype A = yellow, B = red, C = pink, D = dark blue, E = light blue, F = green, G = brown, H = black).

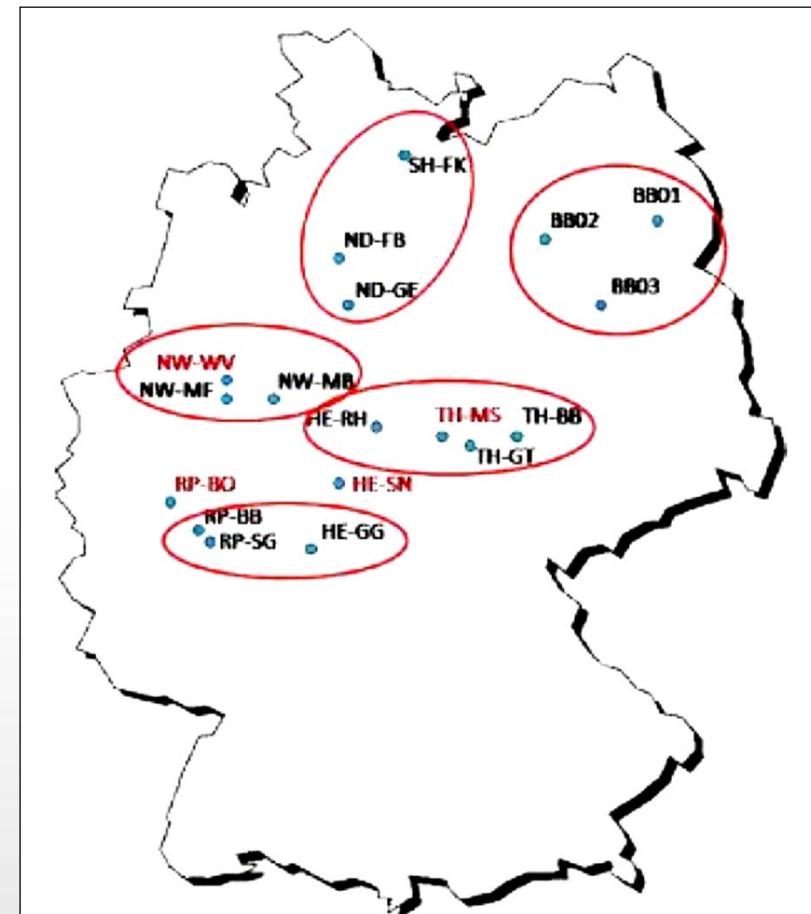
Application of markers for verification of sources

3. Assignment to large-scale regions

Geographic pattern derived from AFLPs,
isozymes and cpDNA-SSR haplotypes

Example *Corylus avellana*

20 populations,
948 individuals



Leinemann L, Hosius B, Steiner W. 2012. Hasel: Genetische Analysen stützen ausgewiesene Vorkommensgebiete - Ergebnisse aus der genetischen Inventur der Haselnuss. TASPO 47, 1

Outlook

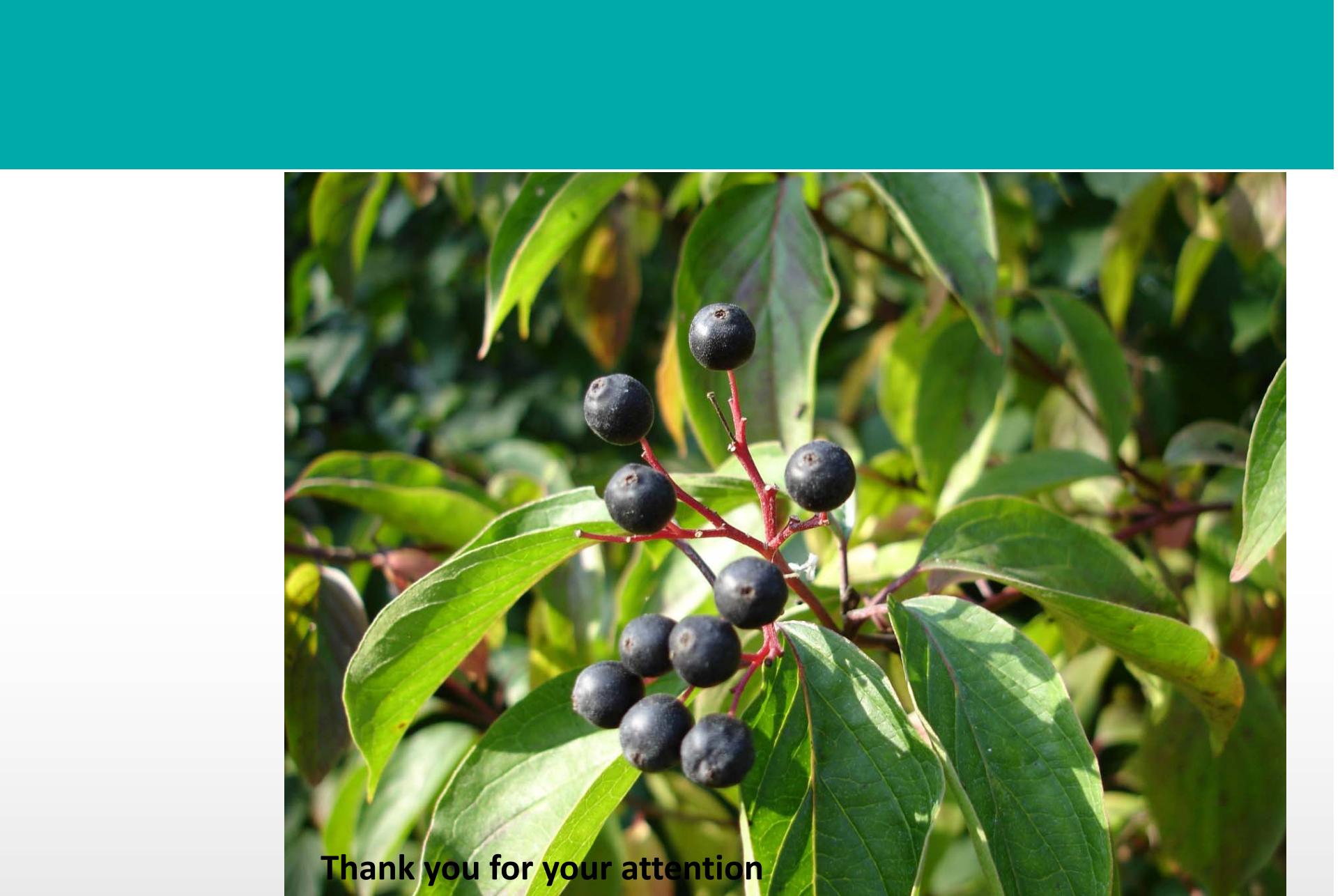
Nuclear and chloroplast markers available

Species name	Deutscher Name
<i>Acer pseudoplatanus</i>	Spitz-Ahorn
<i>Betula pendula</i>	Sand-Birke
<i>Betula pubescens</i>	Moor-Birke
<i>Castanea sativa</i>	Ess-Kastanie
<i>Corylus avellana</i>	Gewöhnliche Hasel
<i>Cytisus scoparius</i>	Besen-Ginster
<i>Fagus sylvatica</i>	Rot-Buche
<i>Frangula alnus</i>	Faulbaum
<i>Fraxinus excelsior</i>	Gewöhnliche Esche
<i>Ligustrum vulgare</i>	Liguster
<i>Populus alba</i>	Silber-Pappel
<i>Populus tremula</i>	Zitter-Pappel
<i>Prunus avium</i>	Vogel-Kirsche
<i>Quercus petraea</i>	Trauben-Eiche
<i>Quercus robur</i>	Stiel-Eiche
<i>Ulmus glabra</i>	Berg-Ulme
<i>Ulmus minor</i>	Feld-Ulme

Markers not yet available

Species name	Deutscher Name
<i>Alnus incana</i>	Grau-Erle
<i>Crataegus laevigata</i>	Zweigriffliger Weißdorn
<i>Euonymus europaea</i>	Pfaffenhütchen
<i>Lonicera nigra</i>	Schwarze Heckenkirsche
<i>Lonicera xylosteum</i>	Rote Heckenkirsche
<i>Rhamnus cathartica</i>	Kreuzdorn
<i>Rosa canina</i>	Hunds-Rose
<i>Rosa majalis</i>	Zimt-Rose
<i>Sambucus racemosa</i>	Trauben-Holunder
<i>Sorbus aucuparia</i>	Eberesche
<i>Tilia platyphyllos</i>	Sommer-Linde
<i>Viburnum lantana</i>	Wolliger Schneeball
<i>Viburnum opulus</i>	Gemeiner Schneeball

Grey: forest tree species under regulations for forest reproductive materials



Thank you for your attention